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AMINO ACIDS SEQUENCE ANALYSIS ON COLLAGEN

Sorana D. Bolboacă ¹, Jäntschi L ²

¹“Tuliu Hatieganu” University of Medicine and Pharmacy Cluj-Napoca, 6 Louis Pasteur, 400349 Cluj-Napoca, Romania, sbolboaca@umfcluj.ro

²Technical University of Cluj-Napoca, 103-105 Muncii Bvd., 400641 Cluj-Napoca, Romania. lori@academicdirect.org

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Abstract. Starting from available information about amino acids properties and sequences on collagen type I chains, the aims of the study were to identify the principal property component and to analyze the similarities within and between collagens on five species. The principal component analysis applied on twenty-four amino acids properties revealed that the hydrophobic or hydrophilic character measured by Wealling et al. is more stable comparing with the other investigated properties. Similarity analysis identified similar and dissimilar within and between studied species from the viewpoint of amino acids sequences on collagen type I alpha 1 and 2 chains.

INTRODUCTION

Collagen, the main protein of connective tissues, is a fibrous protein, inextensible, which can be found at the level of connective tissues from heart, vessels, skin, cornea, cartilage, ligaments, tendons, bone, and teeth. Today are known twenty-eight types of collagens (Veit et al., 2006).

The structure of collagen type I, distributed at the level of all tissues in organism, is known from a long time (Kadler et al., 1996). The COL1A1 and COL1A2 genes which coding the collagens chains have also been identified and characterized: *Homo Sapiens* (Chu et al., 1985; Runyan et al., 2003), *Rattus Norvegicus* (Kwitek et al., 2004; Malfait et al., 2006). Due to its implications in *osteogenesis imperfecta* (Lee et al., 2006), *osteoporosis* (Ralston et al., 2006), *Ehlers-Danlos syndrome* (Pollitt et al., 2006), *Caffey disease* (Gensure et al., 2005), to possible implication in other diseases (e.g. *intracranial aneurysms* - Yoneyama et al., 2004), as indicator of *bone metastasis* - Fukumitsu et al., 2003) the collagen type I is studied by many researchers.

Based on information about amino acids properties and sequences on *collagen type I* chains, the aims of the study were to identify the principal property component and to analyze the similarities within and between collagens on five species.

MATERIAL AND METHOD

Amino Acids Properties

The sample of twenty essential amino acids with twenty-four associated properties has been investigated. The studied properties were: hydrophobic or hydrophilic character measured on different scales (p₁ – Black et al., 1991; p₂ – Kyte and Doolittle, 1982; p₃ – Wimley and White, 1996; p₄ – Hessa et al., 2005; p₅ – Sereda et al., 1994; p₆ – Hoop and Woods, 1981; p₇ – Cornette et al., 1987; p₈ – Eisenberg et al., 1984; p₉ – Rose et al., 1985; p₁₀ – Janin, 1979; p₁₁ – Engelman et al., 1986; p₁₂ – Sweet and Eisenberg, 1983; p₁₄ – Bull and

Breese, 1974; p_{17} – Rosema, 1988; p_{18} – Welling et al., 1985; p_{19} – Parker et al., 1986; p_{20} – Cowan and Whittaker, 1990; p_{21} – Manavalan and Ponnuswamy, 1978; p_{22} – Fauchere and Pliska, 1983; p_{23} – Rao and Argos, 1986; p_{24} – Wilson et al., 1981), partition coefficients (p_{13} – Abraham and Leo, 1987) and energies (p_{15} – Guy, 1985), inter-residue contact energies (p_{16} – Miyazawa and Jernigen, 1985). The data are presented in Table 1.

Table 1

Twenty Amino Acids: Investigated Properties

Name	L	p_1	p_2	p_3	p_4	p_5	p_6	p_7	p_8	p_9	p_{10}	p_{11}	p_{12}	p_{13}	p_{14}	p_{15}	p_{16}	p_{17}	p_{18}	p_{19}	p_{20}	p_{21}	p_{22}	p_{23}	p_{24}
Alanine	A	0.62	1.8	-0.17	0.11	47	-0.50	0.20	0.62	0.74	0.30	1.60	-0.40	0.44	0.61	0.10	5.33	0.39	1.15	2.10	0.35	12.97	0.31	1.36	-0.30
Arginine	R	0.00	-4.5	-0.81	2.58	-26	3.00	1.40	-2.53	0.64	-1.40	-12.3	-0.59	-2.42	0.69	1.91	4.18	-3.95	0.58	4.20	-1.50	11.72	-1.01	0.15	-1.10
Asparagine	N	0.24	-3.5	-0.42	2.05	-41	0.20	-0.50	-0.78	0.63	-0.50	-4.80	-0.92	-1.32	0.89	0.48	3.71	-1.91	-0.77	7.00	-0.99	11.42	-0.60	0.33	-0.20
Aspartate	D	0.03	-3.5	-1.23	3.49	-18	3.00	-3.10	-0.90	0.62	-0.60	-9.20	-1.31	-0.31	0.61	0.78	3.59	-3.81	0.65	10.0	-2.15	10.85	-0.77	0.11	-1.40
Cysteine	C	0.68	2.5	0.24	-0.13	52	-1.00	4.10	0.29	0.91	0.90	2.00	0.17	0.58	0.36	-1.42	7.93	0.25	-1.20	1.40	0.76	14.63	1.54	1.27	6.30
Glutamine	Q	0.25	-3.5	-0.58	2.36	-18	0.20	-2.80	-0.85	0.62	-0.70	-4.10	-0.91	-0.71	0.97	0.95	3.87	-1.30	-0.11	6.00	-0.93	11.76	-0.22	0.33	-0.20
Glutamate	E	0.04	-3.5	-2.02	2.68	8	3.00	-1.80	-0.74	0.62	-0.70	-8.20	-1.22	-0.34	0.51	0.83	3.65	-2.91	-0.71	7.80	-1.95	11.89	-0.64	0.25	0.00
Glycine	G	0.50	-0.4	-0.01	0.74	0	0.00	0.00	0.48	0.72	0.30	1.00	-0.67	0.00	0.81	0.33	4.48	0.00	-1.84	5.70	0.00	12.43	0.00	1.09	1.20
Histidine	H	0.16	-3.2	-0.96	2.06	-42	-0.50	0.50	-0.40	0.78	-0.10	-3.00	-0.64	-0.01	0.69	-0.50	5.10	-0.64	3.12	2.10	-0.65	12.16	0.13	0.68	-1.30
Isoleucine	I	0.94	4.5	0.31	-0.6	100	-1.80	4.80	1.38	0.88	0.70	3.10	1.25	2.46	-1.45	-1.13	8.83	1.82	-2.92	-8.00	1.83	15.67	1.80	1.44	4.30
Leucine	L	0.94	3.8	0.56	-0.55	100	-1.80	5.70	1.06	0.85	0.50	2.80	1.22	2.46	-1.65	-1.18	8.47	1.82	0.75	-9.20	1.80	14.90	1.70	1.47	6.60
Lysine	K	0.28	-3.9	-0.99	2.71	-37	3.00	-3.10	-1.50	0.52	-1.80	-8.80	-0.67	-2.45	0.46	1.40	2.95	-2.77	2.06	5.70	-1.54	11.36	-0.99	0.09	-3.60
Methionine	M	0.74	1.9	0.23	-0.1	74	-1.30	4.20	0.64	0.85	0.40	3.40	1.02	1.10	-0.66	-1.59	8.95	0.96	-3.85	-4.20	1.10	14.39	1.23	1.42	2.50
Phenylalanine	F	1.00	2.8	1.13	-0.32	92	-2.50	4.40	1.19	0.88	0.50	3.70	1.92	2.54	-1.52	-2.12	9.03	2.27	-1.41	-9.20	1.69	14.00	1.79	1.57	7.50
Proline	P	0.71	-1.6	-0.45	2.23	-46	0.00	-2.20	0.12	0.64	-0.30	-0.20	0.49	1.29	-0.17	0.73	3.87	0.99	-0.53	2.10	0.84	11.37	0.72	0.54	2.20
Serine	S	0.36	-0.8	-0.13	0.84	-7	0.30	-0.50	-0.18	0.66	-0.10	0.60	0.55	-0.84	0.42	0.52	4.09	-1.24	-0.26	6.50	-0.63	11.23	-0.04	0.97	-0.60
Threonine	T	0.45	-0.7	-0.14	0.52	13	-0.40	-1.90	-0.05	0.70	-0.20	1.20	-0.28	-0.41	0.29	0.07	4.49	-1.00	-0.45	5.20	-0.27	11.69	0.26	1.08	-2.20
Tryptophan	W	0.88	-0.9	1.85	0.3	84	-3.40	1.00	0.81	0.85	0.30	1.90	0.50	2.56	-1.20	-0.51	7.66	2.13	-1.14	-10.0	1.35	13.93	2.25	1.00	7.90
Tyrosine	Y	0.88	-1.3	0.94	0.68	49	-2.30	3.20	0.26	0.76	-0.40	-0.70	1.67	1.63	-1.43	-0.21	5.89	1.47	0.13	-1.90	0.39	13.42	0.96	0.83	7.10
Valine	V	0.83	4.2	-0.07	-0.31	79	-1.50	4.70	1.08	0.86	0.60	2.60	0.91	1.73	-0.75	-1.27	7.63	1.30	-0.13	-3.70	1.32	15.71	1.22	1.37	5.90

Collagen Type I

The alpha 1 (α_1) and alpha 2 (α_2) chains of collagen type I (CTI) of five species were collected from the Nat. Center for Biotechnology Information [<http://www.ncbi.nlm.nih.gov/>] and have been investigated: *Rattus norvegicus* (Orjel et al., 2006); *Bos taurus* (Fietzek et al., 1975; Shirai et al., 1998); *Danio rerio* (Dubois et al., 2002; Howden, 2007); *Canis lupus* (Lowe et al., 2003); and *Homo sapiens* (Simon et al., 1997; Strausberg et al., 2002).

Method of Analysis

A multivariate analysis on collagen type I amino acids properties were performed by using Principal Component Analysis (PCA) technique. A similarity analysis by using the Oliver algorithm implemented in PHP (Hypertext Preprocessor) was applied on collagen type I chains. The investigation of the minimal number of characters needed to replace, insert or delete in order to transform one amino acid chain into another has also been performed in order to analyze the similarity between and within specie.

RESULTS AND DISCUSSIONS

Principal Component on Amino Acids Properties

The following results were obtained using PCA of amino acids properties:

- ÷ With five exceptions the hydrophobic or hydrophilic character measured on different scales were close to each other in terms of means and standard errors. The exceptions were: p_5 (Sereda et al., 1994), p_{11} (Engelman et al., 1986), p_{19} (Parker et al., 1986), p_{21} (Manavalan and Ponnuswamy, 1978), and p_{24} (Wilson et al., 1981).
- ÷ The correlation matrix on investigated properties identified a very good correlation (≥ 0.95) between the following pairs of hydrophobicity: p_1 (Black et al., 1991) - p_{17} (Rosema,

1988), p₁ (Black et al., 1991) - p₂₀ (Cowan and Whittaker, 1990); and p₁₇ (Rosema, 1988) - p₂₀ (Cowan and Whittaker, 1990).

- ÷ Two variables (hydrophobic or hydrophilic character scales) followed a normal distribution: p₃ (Wimley and White, 1996), and p₁₈ (Welling et al., 1985).
- ÷ A total number of nineteen factors have been identified. The amino acids contribution on factors based on correlation matrix, expressed as extreme values (maximum and minimum) was: p₉ (Rose et al., 1985) with two maximum and one minimum, p₁₃ (Abraham and Leo, 1987) with three minimum, p₁₈ (Welling et al., 1985) with one maximum and three minimum.

Amino Acids on Collagen Type I

The frequency of apparition of amino acids in the alpha 1 and 2 chains according with investigated specie is presented in Table 2.

Table 2

Amino acids distribution in collagen chains

<i>Amino acid (aa)</i>		<i>Homo sapiens</i>		<i>Bos taurus</i>		<i>Canis lupus</i>		<i>Rattus norvegicus</i>		<i>Danio rerio</i>	
Name	Abb	HS_α1	HS_α2	BT_α1	BT_α2	CL_α1	CL_α2	TN_α1	RN_α2	DR_α1	DR_α2
Alanine	A	115	130	143	126	138	123	125	108	162	137
Cysteine	C	10	9	18	9	18	9	0	0	18	8
Aspartate	D	41	43	64	43	64	43	34	23	62	46
Glutamate	E	57	66	76	64	74	65	54	48	82	56
Phenylalanine	F	14	22	24	23	25	21	13	14	28	21
Glycine	G	329	381	389	380	390	381	344	345	382	382
Histidine	H	3	15	9	12	8	12	3	7	10	11
Isoleucine	I	10	32	25	35	26	34	8	19	36	30
Lysine	K	38	50	57	50	56	50	35	21	58	50
Leucine	L	29	61	50	60	47	59	21	34	37	57
Methionine	M	8	10	13	9	15	11	8	5	25	17
Asparagine	N	14	41	29	43	31	42	12	21	35	44
Proline	P	230	231	279	236	278	235	126	113	235	223
Glutamine	Q	30	33	51	36	50	35	25	23	40	37
Arginine	R	51	72	70	73	71	73	52	55	68	72
Serine	S	35	52	58	54	58	52	39	27	62	67
Threonine	T	23	42	44	43	47	51	16	21	53	40
Valine	V	28	55	42	50	42	50	18	39	39	35
Tryptophan	W	1	5	6	5	6	5	0	0	6	5
Tyrosine	Y	3	16	16	13	16	15	5	1	9	14
Unspecified or unknown	X	0	0	0	0	0	0	116	102	0	0
<i>Total</i>		<i>1069</i>	<i>1366</i>	<i>1463</i>	<i>1364</i>	<i>1460</i>	<i>1366</i>	<i>1054</i>	<i>1026</i>	<i>1447</i>	<i>1352</i>

Looking at the data from Table 2 it can be observed that without any exceptions the glycine is the amino acid most abundant in both chains and to all investigated species. Analyzing the amino acids distribution on α1 comparing with α2 within group, statistical significant differences was been identified in two species: *Homo sapiens* (Proline, p = 0.0019), and *Danie rerio* (Glutamate, p = 0.0157). Note that the *Rattus norvegicus* specie is the one with unspecified or unknown amino acids (11% for α1 chain and almost 10% form α2 chains) and without any identified Cysteine or Tryptophan amino acids in sequences of α1 or α2 chain. The strings similarity matrix on collagen chains, expressed as percent of similarity in amino acids within and between species, was calculated (see Table 3). Following are true regarding the string similarities between species: ▪ a good similarity on α1 chain exists

between *Bos taurus* - *Canis lupus*, *Homo sapiens* - *Bos Taurus*, *Bos taurus* - *Danio rerio*, and *Canis lupus*- *Danio rerio*; a good similarity on $\alpha 2$ chain exists between *Bos taurus* - *Canis lupus*, *Homo sapiens* - *Canis lupus*, and *Homo sapiens* - *Bos taurus*.

Table 3.

Collagen chains similarity matrix (expressed as percent)

	BT_α1	BT_α2	CL_α1	CL_α2	DR_α1	DR_α2	HS_α1	HS_α2	RN_α1
BT_α1 = <i>Bos taurus</i> CTIα1	100								
BT_α2 = <i>Bos taurus</i> CTIα2	47.19	100							
CL_α1 = <i>Canis lupus</i> CTIα1	97.43	47.38	100						
CL_α2 = <i>Canis lupus</i> CTIα2	48.71	94.51	50.18	100					
DR_α1 = <i>Danio rerio</i> CTIα1	75.74	49.38	76.37	47.00	100				
DR_α2 = <i>Danio rerio</i> CTIα2	42.77	60.16	43.46	58.13	36.73	100			
HS_α1 = <i>Homo sapiens</i> CTIα1	82.39	35.35	82.56	34.58	62.56	38.08	100		
HS_α2 = <i>Homo sapiens</i> CTIα2	53.23	93.33	52.09	94.51	49.41	66.15	40.49	100	
RN_α1 = <i>Rattus norvegicus</i> CTIα1	72.79	20.93	72.32	20.58	44.22	22.11	74.05	9.34	100
RN_α2 = <i>Rattus norvegicus</i> CTIα2	18.64	70.63	14.00	66.56	25.80	40.62	15.18	66.97	38.94

Table 4.

Collagen chains Levenshtein distances within and between species

	BT_α1	BT_α2	CL_α1	CL_α2	DR_α1	DR_α2	HS_α1	HS_α2	RN_α1
BT_α1	0								
BT_α2	903	0							
CL_α1	70	892	0						
CL_α2	893	98	891	0					
DR_α1	469	902	440	896	0				
DR_α2	909	463	903	492	909	0			
HS_α1	429	882	448	883	731	871	0		
HS_α2	884	115	883	76	895	487	874	0	
RN_α1	1031	921	1033	921	1016	890	650	923	0
RN_α2	1061	941	1059	944	1044	923	671	940	428

The Levenshtein distances, defined as the minimal numbers of characters that must be replace, insert or delete to transform one chain into other are presented in Table 4. With seventy amino acids characters replace, insert or delete the $\alpha 1$ collagen type I chain of *Bos taurus* could be transformed in $\alpha 1$ collagen type I chain of *Canis lupus*. With ninety-eight amino acids characters replace, insert or delete the $\alpha 2$ collagen type I chain of *Bos taurus* could be transformed in $\alpha 2$ collagen type I chain of *Canis lupus*. The greater dissimilarity was identified when collagen type I chains of *Rattus norvegicus* are compared with other species (see Table 3 and 4). This could be explained by the existence of a great number of unknown or unidentified amino acids and/or by the absence of the cysteine and tryptophan (see Table 1).

CONCLUSIONS

The scale of amino acids hydrophobic or hydrophilic is still on debate even if there were proposed many methods. The scale proposed by Welling et al. seems to be more stable on principal component analysis on investigated scales.

The similarity analysis revealed that there are good similarities on Collagen type I amino acids chains between *Bos Taurus* and *Canis lupus*, as well as between *Bos Taurus* and *Homo sapiens*. Due to the lack of amino acids identification on collagen type I of *Rattus norvegicus* the greatest dissimilarities were observed when this specie was compared with the other analyzed species.

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